CONCISE REPORT

Lymphadenopathy-Associated Virus Antibodies and T Cells in Hemophiliacs Treated With Cryoprecipitate or Concentrate

By George F. Gjerset, Gene McGrady, Richard B. Counts, Paul J. Martin, Janine Jason, Susan Kennedy, Bruce Evatt, and John A. Hansen

Evidence for exposure to lymphadenopathy-associated virus (LAV) was investigated in 48 patients with hemophilia, 15 of whom had been treated exclusively with singledonor cryoprecipitate. The prevalence of antibodies to LAV in all patients was 53% in 1983 and 63% in 1984, while in patients treated only with cryoprecipitate, the prevalence was 31% in 1983 and 40% in 1984. Patients treated with any concentrate had a seroprevalence of 65% in 1983 and 77% in 1984. Seropositive patients were more likely to

BNORMALITIES in T cell subsets and T cell function A have been reported in patients with hemophilia A and B.¹⁻⁸ Although the clinical significance of these observations is unclear, hemophiliacs are known to be at high risk for developing the acquired immunodeficiency syndrome (AIDS),⁹ presumably as a consequence of infection with the lymphadenopathy-associated virus (LAV),¹⁰ also known as HTLV III.¹¹ Recent reports have identified antibodies to this virus in otherwise healthy hemophiliacs.^{11,12}

The current study evaluated T cell subsets and T cell function in hemophilic patients receiving commercial concentrates or single-donor cryoprecipitate in relation to the presence or absence of anti-LAV antibodies. Patients were reevaluated after 12 months, allowing us to determine whether there were any changes in LAV antibody prevalence or immune status.

Submitted April 25, 1985; accepted June 5, 1985.

Address reprint requests to Dr George F. Gjerset, Puget Sound Blood Center, Terry at Madison, Seattle, WA 98104.

© 1985 by Grune & Stratton, Inc.

0006-4971/85/6603-0039\$03.00/0

have a significant reduction in the ratio of helper to suppressor T cells, absolute numbers of helper T cells, and T cell function in vitro. Seven of 18 patients who were seronegative in 1983 had seroconverted by 1984. The relative risk of seroconversion for patients using any concentrate since 1981 compared with those using cryoprecipitate only was 3.9 (P = .04). Nevertheless, the rate of conversion in the latter group was 18% per year. • 1985 by Grune & Stratton, Inc.

MATERIALS AND METHODS

Subjects. The study included 48 patients with moderate or severe factor VIII (FVIII) or factor IX (FIX) deficiency treated at the Puget Sound Blood Center, Seattle. Informed consent was obtained according to institutional guidelines. Forty-three were studied in Janaury 1983 and 41 in January 1984. Of 42 patients with FVIII deficiency, 14 were moderately affected (1% to 5% FVIII) and 27 were severely affected (<1% FVIII). The other six patients had severe FIX deficiency. Patients were grouped according to treatment (Table 1). Median age of the patients was 28 years and was comparable for all treatment groups. Controls consisted of healthy volunteers, with a median age of 36 years (range, 13 to 47), with no known risk factors for AIDS.

T cell subsets and function. T helper (T_b) cells and T suppressor (T_s) cells were identified by indirect immunofluorescence using monoclonal antibodies 66.1 to identify the T_h marker, CD4 (T4, Leu-3 equivalent), and 51.1 to identify the T, marker, CD8 (T8 or Leu-2 equivalent).* T cell function was assessed by testing mitogeninduced proliferation of peripheral blood mononuclear cells cultured with 20% heat-inactivated human serum in microtest plates (5 \times 10⁴ cells per 100 µL per well). Phytohemagglutinin (PHA) was added in final concentrations of 0.25, 0.5, and 2.5 μ g/mL, concanavalin A (Con A) in final concentrations of 20, 40, and 80 μ g/mL, and pokeweed mitogen (PWM) in final dilutions of 1:50, 1:125, and 1:200. Cultures stimulated with PHA and Con A were incubated for three days and those with PWM for five days at 37 °C with 5% CO₂. Proliferation was measured as cpm of tritiated thymidine incorporation following a three-hour pulse label.

LAV serology. Anti-LAV antibodies were assayed by Western blot¹⁴ using purified detergent-lysed LAV. LAV-containing supernatants were kindly provided by Dr Luc Montagnier, Institut Pasteur, Paris.¹⁰ Seropositivity was defined as a reaction with any of the viral antigens p18, p25, or p41.

Table 1. Patient Characteristics

Tγpe of Therapγ●	No. of Patients†	Amount of Replacement Therapy‡	
		Cryoprecipitate	Concentrate
I. Cryoprecipitate only	15 (8)	57 (8–193)	0
II. Cryoprecipitate + concentrate	19 (13)	104 (27–252)	1 (<1-68)
III. Concentrate only	14 (13)	0	127 (39-323)
Total	48		

*Cryoprecipitate was prepared from single volunteer donor units.¹³ FVIII or FIX concentrate was obtained commercially.

+Five patients included in the 1984 evaluation were not studied in 1983, and seven patients studied in 1983 were not available in 1984. Numbers in parentheses indicate numbers of patients severely affected.

* Numbers represent units × 10³ per year. One donor unit of cryoprecipitate is equivalent to 100 FVIII units; one bottle of concentrate is equivalent to 1,000 FVIII or FIX units.

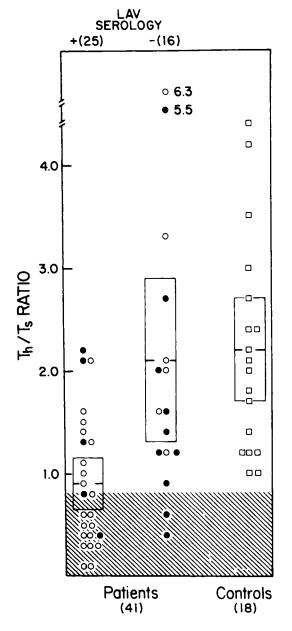
From the Puget Sound Blood Center, Fred Hutchinson Cancer Research Center, and the University of Washington, Seattle; and the Division of Host Factors, Centers for Infectious Disease and for Disease Control, Public Health Service, Department of Health and Human Services (DHHS), Atlanta.

Supported in part by DHHS grants No. HL17265, CA29548, CA18020, and CA15704. G.F.G. was supported by the National Research Service Training Grant.

Table 2. Anti-LAV Antibody in Patients With Hemophilia

Type of Replacement Therapy	Positive LAV Serology		Rate of
	1983	1984	Seroconversion*
I. Cryoprecipitate only	5/16 (31%)	6/15 (40%)	2/10 (20%)
II. Cryoprecipitate + concentrate	10/15 (67%)	10/14 (71%)]	5/8 (62%)
III. Concentrate only	7/11 (64%)	10/12 (83%) }	
Total	22/42 (53%)	26/41 (63%)	7/18 (39%)

*Eighteen patients evaluated in both 1983 and 1984 were seronegative in 1983.



RESULTS

LAV serology. Antibodies to LAV were detected in 53% of patients in 1983 and 63% in 1984 (Table 2). Seroprevalence was significantly greater in patients treated with concentrate (65% in 1983, 77% in 1984) compared to patients treated only with cryoprecipitate (31% in 1983, 40% in 1984) (P < .02, chi-square test). Of the 36 patients evaluated in both 1983 and 1984, 18 were seronegative in 1983. Ten of the 18 seronegative patients had received only cryoprecipitate; an additional seronegative patient received only cryoprecipitate since January 1981. Of these 11 patients, two (18%) seroconverted. Thus, the relative risk of seroconversion for patients using any concentrate since January 1981 was 3.9 (P = .04, Fisher's exact test).

Relationship between LAV serology and T cell subsets. T_h/T_s ratios were significantly lower in seropositive patients compared to seronegative patients in 1984 (median, 0.8 v 1.6; P < .002, Mann-Whitney U test) (Fig 1). A similar association between LAV serology and T_h/T_s ratio was also

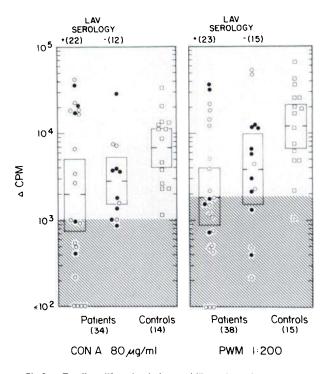


Fig 1. T_h/T_s ratios in hemophilic patients according to presence or absence of anti-LAV antibody (\bullet , patients receiving cryoprecipitate only; \circ , patients receiving any concentrate; \Box , controls). Means and 95% confidence intervals are indicated by windows. The shaded area represents the abnormally low range as determined by testing healthy normal controls (n = 60).

Fig 2. T cell proliferation in hemophilic patients in response to stimulation with Con A (80 μ g/mL) or PWM (final dilution, 1:200). Shaded area represents the abnormally low range of response as determined by testing healthy normal controls (mean -2 SD).

GJERSET ET AL

observed in 1983 (data not shown). In the seven patients who seroconverted between 1983 and 1984, however, no significant change was seen in the T_h/T_s ratio. The T_h/T_s ratio also remained stable in 19 patients who were already seropositive in 1983.

Relationship between LAV serology and T cell function. Responses to PWM, but not Con A, were significantly lower in seropositive patients than controls (P = .005, Mann-Whitney U test) (Fig 2). Low responses to PWM or Con A (< control mean - 2 SD) were significantly more frequent in seropositive patients than controls (odds ratio >21, $\chi^2 > 10.1$; P < .01). Responses in seronegative patients, however, did not differ significantly from those in seropositive patients or in controls. When patients were stratified according to type of replacement treatment, no significant association was seen between LAV serology and T cell response.

Clinical status. Thirty-six of 48 patients first evaluated in January 1983 were available for evaluation in January 1984, and all but four remained well. One 10-year-old seropositive patient in group II developed molluscum contagiosum, eosinophilia, and intermittent abdominal pain. A 35-year-old seropositive patient in group IV had immune thrombocytopenia requiring splenectomy. Two patients died, one after trauma and the other with a ruptured aneurysm.

DISCUSSION

Antibodies to LAV were detected in hemophilic patients treated only with cryoprecipitate (40% in 1984) as well as in patients treated with concentrate (77% in 1984). The rate of seroconversion between 1983 and 1984 was greater for patients receiving any concentrate (18% v 71%) since Jan-

1. Lederman MM, Ratnoff OD, Scillian JJ, Jones PK, Schacter B: Impaired cell-mediated immunity in patients with classic hemophilia. N Engl J Med 308:79, 1983

2. Menitove JE, Aster RH, Casper JT, Lanier SJ, Gottschall JL, Williams JE, Bill JC, Wheeler DV, Piaskowski V, Kirchner P, Montgomery RR: T-lymphocyte subpopulations in patients with classic hemophilia treated with cryoprecipitate and lyophilized concentrates. N Engl J Med 308:83, 1983

3. Luban NLC, Kelleher Jr JF, Reaman GH: Altered distribution of T-lymphocyte subpopulations in children and adolescents with haemophilia. Lancet 1:503, 1983

4. Kaplan J, Sarnaik S, Gitlin J, Lusher J: Diminished helper/ suppressor lymphocyte ratios and natural killer activity in recipients of repeated transfusion. Blood 64:308, 1984

5. Weintrub PS, Koerper MA, Addiego JE, Drew WL, Lennette ET, Miner R, Cowan MJ, Ammann AJ: Immunologic abnormalities in patients with hemophilia A. J Pediatr 103:692, 1983

6. de Schazo RD, Andes WA, Nordberg J, Newton J, Paul C, Bozelka B: An immunologic evaluation of hemophiliac patients and their wives. Ann Intern Med 99:159, 1983

7. Landay A, Poon MC, Abo T, Stagno S, Lurie A, Cooper MD: Immunologic studies in asymptomatic hemophilia patients: Relationship to acquired immune deficiency syndrome (AIDS). J Clin Invest 71:1500, 1983

8. Gjerset FG, Martin PJ, Counts RB, Fast LD, Hansen JA: Immunologic status of hemophilia patients treated with cryoprecipitate or lyophilized concentrate. Blood 64:715, 1984 uary 1981. Thus, although the prevalence of antibody and the relative risk of seroconversion (3.9) was greater in the concentrate group, these data also indicate that the transfusion of large amounts of cryoprecipitate collected from healthy volunteer donors but not screened for anti-LAV antibodies can be associated with substantial risk of exposure to infectious virus.

A significant association was seen between anti-LAV antibodies and abnormalities in the T_h/T_s ratios and T cell function, suggesting that alterations in T cell subsets and T cell function can be related in transfusion-related exposure to LAV. However, abnormal T_h/T_s ratios and T cell responses were not detected in all patients exposed to LAV, and progressive changes were not observed in patients who seroconverted during the course of study. With the exception of two individuals, patients also remained well and free of unusual infections or other clincal evidence of immunodeficiency.

Whether the immune status of these patients will continue to be stable, even assuming no further exposure to LAV, is unknown. Screening blood donors for anti-LAV antibodies will presumably decrease the risk of AIDS transmission. Additional measures, however, such as heat inactivation,¹⁵ continue to represent important alternatives for preventing infection in patients exposed to blood products from a large number of donors.

ACKNOWLEDGMENT

We thank Dr J. Steve McDougal, Centers for Disease Control, for his efforts in making the Western blot studies possible and Maribel Clements, RN, Clinical Associate, Puget Sound Blood Center Hemophilia Program, for her work in coordinating this study.

REFERENCES

9. Centers for Disease Control: Update: Acquired immunodeficiency syndrome (AIDS) in persons with hemophilia. MMWR 33:589, 1984

10. Vilmer E, Rouzioux C, Vesinet Brun F, Fischer A, Chermann JC, Barre-Sinoussi F, Gazengel C, Daugnet C, Manigne P, Griscelli C, Montagnier L: Isolation of a new lymphotropic retrovirus from two siblings with hemophilia, one with AIDS. Lancet 1:573, 1984

11. Schupbach J, Popovic M, Gilden RV, Gond MA, Sarngadharan MG, Gallo RC: Serological analysis of a subgroup of human T lymphotropic retroviruses (HTLV-III) associated with AIDS. Science 224:503, 1984

12. Melbye M, Froebel KS, Madhok R, Biggar RJ, Sarin PS, Stenbjerg S, Lowe GDO, Forbes CD, Goedert JJ, Gallo RC, Ebbesen P: HTLV-III seropositivity in European haemophiliacs exposed to factor VIII concentrate imported from the U.S.A. Lancet 2:1444, 1984

13. Slichter SJ, Counts RB, Henderson R, Harker LA: Preparation of cryoprecipitated factor VIII concentrates. Transfusion 16:616, 1976

14. Tsang VC, Peralta JM, Simons AR: Enzyme-linked immunoelectrotransfer blot techniques (EITB) for studying the specificities of antigens and antibodies separated by gel electrophoresis. Methods Enzymol 92:377, 1983

15. Levy LA, Mitra G, Mozen MM: Recovery and inactivation of infectious retroviruses from factor VIII concentrates. Lancet 2:722, 1984



Lymphadenopathy-associated virus antibodies and T cells in hemophiliacs treated with cryoprecipitate or concentrate

GF Gjerset, G McGrady, RB Counts, PJ Martin, J Jason, S Kennedy, B Evatt and JA Hansen

Updated information and services can be found at: http://www.bloodjournal.org/content/66/3/718.full.html Articles on similar topics can be found in the following Blood collections

Information about reproducing this article in parts or in its entirety may be found online at: http://www.bloodjournal.org/site/misc/rights.xhtml#repub_requests

Information about ordering reprints may be found online at: http://www.bloodjournal.org/site/misc/rights.xhtml#reprints

Information about subscriptions and ASH membership may be found online at: http://www.bloodjournal.org/site/subscriptions/index.xhtml